

Amendments to the Specification

Please replace paragraph number 0128 with the following rewritten paragraph:

Two of the resulting probes (TM63 and TM74), shown in Table 1, below, were labeled, mixed, and used to screen the above genomic library. Oligos were labeled with $\gamma^{32}\text{P}$ ATP using T4 polynucleotide kinase as described (Ausubel, *et al*, eds, 1994. "Current Protocols in Molecular Biology," John Wiley and Sons, Inc.,) and cleaned up using Elutipis (Schleicher & Schuell). Hybridization of duplicate filters was carried out in a Bellco hybridization oven at 37°C using the SSPE protocol as described (Ausubel, *et al*, eds, "Current Protocols in Molecular Biology," John Wiley and Sons, Inc., 1994). Filters were washed in 6X SSC with 0.5%SDS (Ausubel, *et al*, eds, "Current Protocols in Molecular Biology," John Wiley and Sons, Inc., 1994) at 37°C. Filters were then washed at successively higher temperatures in 3 M TMAC (Ausubel, *et al*, eds, "Current Protocols in Molecular Biology," John Wiley and Sons, Inc., 1994) until very little radioactivity could be detected with a survey meter (generally 45 - 55°C). Upon exposure to X-Ray film (Kodak X-Omat), colonies which were evident on both replicate filters were picked with a wooden toothpick and transferred to a fresh nylon filter overlaid onto an LB/ampicillin plate. This procedure was repeated until a homogeneous population was achieved.

Table 1: oligonucleotides (SEQ ID NOS: 13-31, respectively, in order of appearance) with DNA sequence and approximate coordinates relative to the ATG start codon on SEQ ID NOS: 7 or 12.

| <u>Name</u> | <u>Length</u> | <u>Sequence (5' to 3')</u> | <u>Coordinates</u> |
|-------------|---------------|--------------------------------|--------------------|
| TM63 | 30 | CGCGTTCAGGACGCATACTCCGTTCGCTGC | 838-867 |
| TM74 | 24 | GCCCATGGAAACGTGGTCTTCCTG | 1370-1393 |
| TM85 | 21 | ATCATCATGCCCGAGTCCACA | 1156-1176 |
| TM87 | 21 | GCCATCAGGAAGACCACGTTT | 990-971 |

| | | | |
|-------|----|---------------------------------------|-------------------|
| TM89 | 20 | ATGCAGGAAGACCACGTTTC | 1246-1265 |
| TM91 | 21 | ATCGAGGTCCGCCAATGCCAT | 648-628 |
| TM92 | 18 | ACCGGAGCAGCCCAGTGA | 441-424 |
| TM93 | 20 | TGCTTGAAGTATTGCGCCAG | 1403-1422 |
| TM94 | 18 | GATCCTCGGGTGCGATGT | 226-209 |
| TM95 | 18 | ATGCTGATCGGGCTTCGT | 92-74 |
| TM96 | 27 | ATTTGATT <u>CATATGG</u> CTTCCGCTCCTC | -11- +16 |
| TM97 | 28 | ATCTT <u>GGATCC</u> GAACATGGTGCGTTGCA | Beyond C-Terminus |
| TM98 | 18 | AGCACCAGAT CGATGCAC | 128-145 |
| TM99 | 18 | TGGCATGGGTGAACCGGT | 267-284 |
| TM101 | 18 | ATCAGCGTTGAAGCCCAG | 682-699 |
| TM103 | 18 | ACGTGCTGGACTTCCTTG | 1019-1036 |
| TM105 | 18 | GTGCATAAGGCCCTCGAA | 1501-1518 |
| TM106 | 18 | GAGCTTCGAGGGCCTTAT | 1522-1505 |
| TM109 | 18 | CGAGCAACGCAGCGAGTA | 870-853 |

Please replace paragraph number 146, with the following rewritten paragraph:

Purified HAL has been determined to have approximately 40 I.U./mg of activity at 37°C. The temperature optimum was found to be 45°C (figure 7). The graph shows that the enzyme maintains a significant level of activity at physiological temperature conditions. ~~Also below is a~~

~~graph depicting the effect of pH on HAL activity.~~ The activity profile of HAL at various pH is depicted in figure 8. The enzyme is active over a wide range of pH, with highest activity around pH 8.2 and high activity in physiological conditions.